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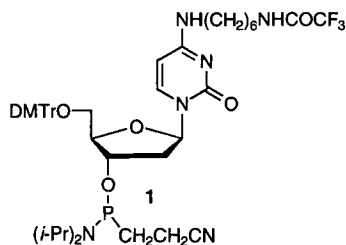
A SIMPLE SYNTHESIS OF *N*⁴-(6-AMINOHEXYL)-2'-DEOXY-5'-*O*-(4,4'-DIMETHOXYTRITYL)CYTIDINE

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Abstract: The title compound was synthesized by a transamination reaction between *N*⁴-benzoyl-2'-deoxy-5'-*O*-(4,4'-dimethoxytrityl)cytidine and hexane-1,6-diamine in the presence of 1,5,7-triazabicyclo(4.4.0)dec-5-ene (TBD).

Oligonucleotides labeled with lanthanide chelates are routinely used as tools in research and diagnostic applications as probes for detection of specific nucleic acid sequences. For example, in mixed-phase hybridization assays up to 20 aminohexane-modified deoxycytidine phosphoramidites (**1**) are coupled to the 5'-end of oligonucleotides.^{1,2} After deprotection of the modified oligonucleotide, the primary amino functions are labeled with a photoluminescent europium chelate.



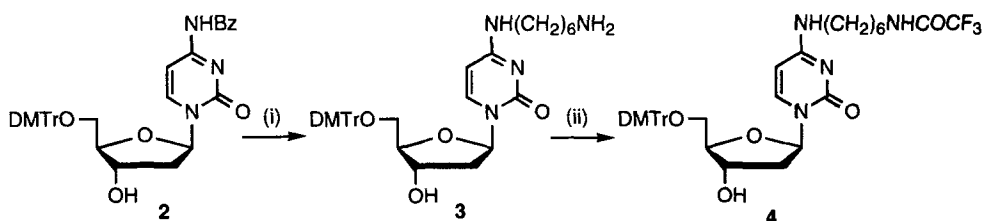
According to the original synthetic strategy,^{3,4} the phosphoramidite **1** has been prepared as follows: the hydroxyl groups of 2'-deoxycytidine were protected using 1,3-dichloro-1,1,3,3-tetraisopropylidisiloxane and the exocyclic amino function was tosylated. Substitution of the tosylamido group with hexane-1,6-diamine introduced the desired tether. Protection of the linker amino function, desilylation, dimethoxytritylation and phosphitylation completed the reaction sequence. Since several synthetic steps and the use of the expensive 1,3-dichloro-1,1,3,3-tetraisopropylidisiloxane to form a transient hydroxyl protecting group are needed, the price of scaling up the experiment is unreasonably high.

Several other methods for cytosine tethering have been reported. For example, Roget *et al.*⁵ obtained *N*⁴-(6-aminohexyl)-2'-deoxy-5'-*O*-(4,4'-dimethoxytrityl)cytidine (**3**) in good yield by allowing 2'-deoxy-5'-*O*-(4,4'-dimethoxytrityl)-4-thiouridine to react with hexane-1,6-diamine. The 4-thio derivative was, in turn, synthesized in four steps from 2'-deoxyuridine.⁶ An alternative approach for tethering involves bisulfite-ion-catalyzed transamination of 2'-deoxycytidine with alkane- α,ω -diamines.⁷ The major drawback of this method is the laborious purification procedure of *N*⁴-(ω -alkylamino)-2'-deoxycytidine.⁸

It is well established that transamination of *N*⁴-benzoylated cytosine residues is a serious side reaction when protected oligonucleotides are treated with aqueous primary amines.⁹⁻¹⁴ Depending on the nature of the amine and the reaction conditions, the transamination has been reported to occur in up to 40% yield.¹³ It is also known that organic solvents and strong bases, such as 1,5,7-triazabicyclo(4.4.0)dec-5-ene (TBD), have an enhancing effect on the transamination reaction.¹⁴ In the present case these observations were exploited in the preparation of the 2'-deoxycytidine derivative **3** (Scheme). Accordingly, treatment of commercially available or easily accessible¹⁵ *N*⁴-benzoyl-2'-deoxy-5'-*O*-(4,4'-dimethoxytrityl)cytidine (**2**) with hexane-1,6-diamine in propan-2-ol in the presence of TBD gave rise to **3**. About 20% of 2'-deoxy-5'-*O*-(4,4'-dimethoxytrityl)cytidine, the product of debenzoylation, was formed. The desired product **3** was easily isolated from the reaction mixture in 56% yield by extraction followed by silica gel column chromatography. After protection of the primary amino function of **3** as its trifluoroacetamide, the nucleoside **4** was ready for phosphitylation and incorporation into oligonucleotides.

EXPERIMENTAL

General. *N*⁴-Benzoyl-2'-deoxy-5'-*O*-(4,4'-dimethoxytrityl)cytidine (**2**) was either purchased from Sigma or prepared in one pot from 2'-deoxycytidine according to Jones.¹⁵ Methyl trifluoroacetate was prepared by mixing trifluoroacetic acid and anhydrous methanol and collecting the product boiling at 43 °C. No catalyst or external heating was used. Adsorption column chromatography was performed on columns packed with silica gel 60 (Merck). Analytical TLC was conducted on silica gel 60 F₂₅₄ plates (Merck). The



Scheme. (i) Hexane-1,6-diamine and TBD in propan-2-ol, overnight at 60 °C. (ii) Methyl trifluoroacetate in dichloromethane, 1h at rt.

following solvent systems were used as the eluents: *A*: CH₂Cl₂:MeOH 9:1 (v/v); *B*: CH₂Cl₂:MeOH:Et₃N 7:2:1 (v/v/v). NMR spectra were recorded on a Jeol LA-400 spectrometer operating at 399.8 and 376.0 MHz for ¹H and ¹⁹F, respectively. The signal of Me₄Si was used as an internal (¹H) and trifluoroacetic acid (¹⁹F) as an external reference. Coupling constants are given in Hertz. Mass spectra were recorded on a VG ZabSpec-aoTOF instrument (FAB⁺).

***N*⁴-(6-Aminohexyl)-2'-deoxy-5'-*O*-(4,4'-dimethoxytrityl)cytidine (3).** Compound **2** (2.0 g, 3.16 mmol) was dissolved in propan-2-ol (10 mL). Hexane-1,6-diamine (5 g) and TBD (1.3g) were added, and the mixture was stirred overnight at 60 °C. The work up was performed as described by Roget *et al.*⁵ with some modifications. Accordingly, the reaction mixture was cooled to room temperature and the solvent was evaporated off *in vacuo*. The residue was dissolved in chloroform (100 mL), extracted with 0.1M NaOH (2x50 mL) and water (5x50 mL). The organic phase was dried (MgSO₄) and concentrated. The residue was dissolved dichloromethane and applied onto a silica gel column. The column was eluted first with *eluent A* to remove yellow impurities and 2'-deoxy-5'-*O*-(4,4'-dimethoxytrityl)cytidine, and then with *eluent B* to elute the product. Pure fractions were pooled and concentrated to give the title compound as a white foam (1.11 g, 56%). *R*_f(*A*): 0.0; *R*_f(*B*): 0.40. Its ¹H NMR spectrum was in agreement with that reported in the literature.⁵ MS: 629 [M⁺+H].

2'-Deoxy-5'-*O*-(4,4'-dimethoxytrityl)-*N*⁴-(6-*N*-trifluoroacetamidohexyl)cytidine (4). Compound **3** (1.5 g, 2.4 mmol) was dissolved in dry dichloromethane (10 mL). Freshly

distilled methyl trifluoroacetate (1 mL) was added and the mixture was stirred for 1h at room temperature. All volatile materials were removed *in vacuo*. Purification on silica gel column chromatography (*eluent A*) yielded the title compound as a white foam (1.56 g, 90%). $R_f(A)$ 0.49. ^1H NMR (CDCl_3): δ 7.78 (1H, d, J 7.9, H-6), 7.76 (1H, br, $\text{N}^4\text{-H}$); 7.33-7.25 (9H, DMTr), 7.21 (1H, br t, NHCOCF_3), 6.83 (4H, d, J 8.9, DMTr), 6.32 (1H, t, $J_{1',2'}$ and $J_{1',2''}$ 6.0, H-1'), 5.36 (1H, d, J 7.9, H-5), 4.45 (1H, m, H-3'), 4.00 (1H, m, H-4'), 3.78 (6H, s, $2\times\text{OCH}_3$), 3.47 (2H, dd, $J_{4',5'}$ 3.6 and $J_{5',5''}$ 10.6, H-5'), 3.39 (1H, dd, $J_{4',5'}$ 3.8 and $J_{5',5''}$ 10.6, H-5''), 3.34 (4H, m, NHCH_2 and $\text{CH}_2\text{NHCOCF}_3$), 2.52 (1H, m, H-2'), 2.19 (1H, m, H-2''), 1.59 (4H, m, $2\times\text{CH}_2$), 1.41 (4H, m, $2\times\text{CH}_2$). ^{19}F NMR (CDCl_3): δ -76.56. HRMS Found: 725.3159 [$\text{M}^+ + \text{H}$]. Calcd. for $\text{C}_{38}\text{H}_{44}\text{F}_3\text{N}_4\text{O}_7$: 725.3162.

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